

# Package: drugsens (via r-universe)

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**Title** Automated Analysis of 'QuPath' Output Data and Metadata Extraction

**Description** A comprehensive toolkit for analyzing microscopy data output from 'QuPath' software. Provides functionality for automated data processing, metadata extraction, and statistical analysis of imaging results. The methodology implemented in this package is based on Labrosse et al. (2024) [doi:10.1016/j.xpro.2024.103274](https://doi.org/10.1016/j.xpro.2024.103274) "Protocol for quantifying drug sensitivity in 3D patient-derived ovarian cancer models", which describes the complete workflow for drug sensitivity analysis in patient-derived cancer models.

**Version** 0.1.0

**BugReports** <https://git.scicore.unibas.ch/ovca-research/drugsens/-/issues>

**SystemRequirements** QuPath™ 4.0.0 or higher

**URL** <https://git.scicore.unibas.ch/ovca-research/drugsens/>

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**License** MIT + file LICENSE

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change\_data\_format\_to\_longer  
*Data format changer*

---

### Description

This function gets the count data dataframe, that has a wider format and it returns a longer-formatted dataframe

### Usage

```
change_data_format_to_longer(
  .data,
  pattern_column_markers = "_ratio_of_total_cells",
  unique_name_row_identifier = "filter_image",
  additional_columns = TRUE
)
```

### Arguments

`.data` The markers count dataframe that is coming from the processing of the microscopy data

`pattern_column_markers` The markers' pattern name to obtain the column with ratios of the markers (it defaults to "\_ratio\_of\_total\_cells")

`unique_name_row_identifier` String that indicates the unique identifier for each image, defaults as "filter\_image"

`additional_columns` columns that can be additionally added to the longer formatted dataframe, "Defaults as c("Treatment", "PID", "Image\_number", "Tissue", "Concentration", "DOC")"

### Details

Reformat the counts data in longer format

**Value**

A dataframe/tibble.

**Examples**

```
# Set up relabeling list
list_of_relabeling <- list(
  "PathCellObject" = "onlyDAPIPositive",
  "cCasp3" = "cCASP3",
  "E-Cadherin: cCASP3" = "E-Cadherin and cCASP3",
  "EpCAM_E-Cadherin" = "E-Cadherin",
  "EpCAM_E-Cadherin and cCASP3" = "E-Cadherin and cCASP3"
)

# Load and process example data
bind_data <- data_binding(
  path_to_the_projects_folder = system.file("extdata/to_merge/", package = "drugsens")
)
counts_dataframe <- make_count_dataframe(bind_data)

# Convert to long format
plotting_ready_dataframe <- change_data_format_to_longer(counts_dataframe)
```

---

data\_binding

*Merge all the dataframes coming out from the QuPath*

---

**Description**

This function identifies string patterns in the dataset, fills the dataframe with that information, and combines all data into a single file

**Usage**

```
data_binding(
  path_to_the_projects_folder,
  files_extension_to_look_for = "csv",
  recursive_search = FALSE,
  forcePath = NULL
)
```

**Arguments**

path\_to\_the\_projects\_folder  
String/Path The path where the files coming out of QuPath are located

files\_extension\_to\_look\_for  
String The extension of the file outputted from QuPath, (default is "csv")

recursive\_search      Boolean, it defined the behavior of the file search, if recursive or not, (default is FALSE)  
forcePath              String defining an alternative path to the config file

**Value**

A concatenated dataframe from all the files within the indicated path

**Examples**

```
## Not run:  
bind_data <- data_binding(path_to_the_projects_folder = system.file("extdata/to_merge/",  
                           package = "drugsens"))  
  
## End(Not run)
```

---

generate\_qupath\_script

*Generate the groovy script used for the analysis*

---

**Description**

Generate a useful script to consistently save the output data from QuPath in .csv format following the naming conventions followed during the package development.

**Usage**

```
generate_qupath_script(output_dir = NULL)
```

**Arguments**

output\_dir      Directory where the script should be saved. If NULL, uses tempdir()

**Value**

Invisibly returns the path to the generated script file.

**Examples**

```
## Not run:  
# Generate script in a temporary directory  
generate_qupath_script()  
  
# Generate script in a specific directory  
output_dir <- tempdir()  
generate_qupath_script(output_dir = output_dir)  
  
## End(Not run)
```

---

`get_QC_plots`*Plot some QC plots to define that everything ran correctly*

---

### Description

Plot data to visualize immediate trends. This function expects data that has been processed through `make_count_dataframe()` and `change_data_format_to_longer()` to ensure the correct data structure for plotting.

### Usage

```
get_QC_plots(  
  .data,  
  patient_column_name = "PID",  
  colors = c("darkgreen", "red", "orange", "pink"),  
  save_plots = FALSE,  
  folder_name = NULL,  
  x_plot_var = "Treatment_complete",  
  isolate_a_specific_patient = NULL  
)
```

### Arguments

<code>.data</code>	The preprocessed data (after running <code>make_count_dataframe()</code> and <code>change_data_format_to_longer()</code> ) merged data.frame that should be visualized
<code>patient_column_name</code>	The PID's column name in the merged data.frame (defaults to "PID")
<code>colors</code>	A list of colors to supply to personalize the plot, defaults to <code>c("darkgreen", "red", "orange", "pink")</code>
<code>save_plots</code>	A Boolean value indicating if the plots should be saved or not (default is FALSE)
<code>folder_name</code>	A string indicating the name of the folder where to save the plots if <code>save_plots</code> is TRUE
<code>x_plot_var</code>	A string indicating the treatment's full name for the QC plots (default is "Treatment_complete")
<code>isolate_a_specific_patient</code>	A string indicating the patient name to isolate for single plot case (default is NULL)

### Value

Invisibly returns NULL, but saves plots to disk if `save_plots` is TRUE

**Examples**

```

# First process example data
example_path <- system.file("extdata/to_merge/", package = "drugsens")
raw_data <- data_binding(path_to_the_projects_folder = example_path)
count_data <- make_count_dataframe(raw_data)
processed_data <- change_data_format_to_longer(count_data)

# Create and save plots to temporary directory
temp_dir <- file.path(tempdir(), "qc_plots")
get_QC_plots(
  processed_data,
  save_plots = TRUE,
  folder_name = temp_dir
)

# Create plots for a specific patient
get_QC_plots(
  processed_data,
  isolate_a_specific_patient = "B39",
  save_plots = TRUE,
  folder_name = temp_dir
)

```

---

```
get_QC_plots_parsed_merged_data
```

*Plot QC plots and calculate statistics for bound data*

---

**Description**

This function creates quality control plots and calculates basic statistics for microscopy data. The plots provide visual insights into marker expression patterns and data quality.

**Usage**

```

get_QC_plots_parsed_merged_data(
  .data,
  list_of_columns_to_plot = NULL,
  save_plots = FALSE,
  saving_plots_folder = NULL,
  save_plots_in_patient_specific_subfolders = TRUE,
  fill_color_variable = NULL,
  PID_column_name = "PID",
  isolate_specific_drug = NULL,
  isolate_specific_patient = NULL,
  drug_column_name = "Treatment",
  save_list_of_plots = TRUE,
  p_height = 10,

```

```

    p_width = 10,
    verbose = TRUE
  )

```

### Arguments

<code>.data</code>	The preprocessed data frame to analyze
<code>list_of_columns_to_plot</code>	Columns to include in plots. If NULL, all numeric columns are used.
<code>save_plots</code>	Logical, whether to save plots to files. Defaults to FALSE.
<code>saving_plots_folder</code>	Directory for saving plots. If NULL and <code>save_plots=TRUE</code> , uses a subdirectory of <code>tempdir()</code> .
<code>save_plots_in_patient_specific_subfolders</code>	Logical, whether to create patient subdirectories. Defaults to TRUE.
<code>fill_color_variable</code>	Variable name for plot color filling
<code>PID_column_name</code>	Column name for patient IDs. Defaults to "PID".
<code>isolate_specific_drug</code>	Drug name to subset data
<code>isolate_specific_patient</code>	Patient ID to subset data
<code>drug_column_name</code>	Column name for drug information. Defaults to "Treatment".
<code>save_list_of_plots</code>	Logical, whether to return list of plot objects. Defaults to TRUE.
<code>p_height</code>	Plot height in inches. Defaults to 10.
<code>p_width</code>	Plot width in inches. Defaults to 10.
<code>verbose</code>	Logical, whether to show progress messages. Defaults to TRUE.

### Value

If `save_list_of_plots=TRUE`, returns a named list of ggplot objects. Otherwise returns invisible(NULL).

### Examples

```

# First load and process example data
example_path <- system.file("extdata/to_merge/", package = "drugsens")
raw_data <- data_binding(path_to_the_projects_folder = example_path)
count_data <- make_count_dataframe(raw_data)
processed_data <- change_data_format_to_longer(count_data)

# Basic usage - create plots for all patients
plots <- get_QC_plots_parsed_merged_data(processed_data)

```

```

# Save plots to a temporary directory
temp_dir <- file.path(tempdir(), "qc_plots")
plots <- get_QC_plots_parsed_merged_data(
  processed_data,
  save_plots = TRUE,
  saving_plots_folder = temp_dir
)

# Focus on a specific patient
plots <- get_QC_plots_parsed_merged_data(
  processed_data,
  isolate_specific_patient = "B39"
)

# Color plots by tissue type
plots <- get_QC_plots_parsed_merged_data(
  processed_data,
  fill_color_variable = "Tissue"
)

```

---

make\_count\_dataframe *Count the main marker expression*

---

## Description

This function counts every single marker present in the "Name" column of the data.frame and return a dataframe of the counts per marker

## Usage

```

make_count_dataframe(
  .data,
  unique_name_row_identifiser = "filter_image",
  name_of_the_markers_column = "Name"
)

```

## Arguments

`.data` The dataframe that is coming from the processing of the microscopy data

`unique_name_row_identifiser` The name of the column of the `.data` where the unique name can be used to counts (it defaults to "filter\_image")

`name_of_the_markers_column` The name of the column of the `.data` where the marker names are expressed (ie E-Caderin, DAPI), "Defaults as Name"



**Value**

A dataframe/tibble.

**Examples**

```
# First load example data
pkg_path <- system.file("extdata/to_merge/", package = "drugsens")
bind_data <- data_binding(
  path_to_the_projects_folder = pkg_path,
  files_extension_to_look_for = "csv"
)

# Process the data
counts_dataframe <- make_count_dataframe(bind_data)

# Convert to plotting format
plotting_ready_dataframe <- change_data_format_to_longer(counts_dataframe)

# Example with custom parameters
make_count_dataframe(
  bind_data,
  name_of_the_markers_column = "Name",
  unique_name_row_identifier = "filter_image"
)
```

---

make_run_config	<i>Generates and use a config txt file</i>
-----------------	--

---

**Description**

When this function run the first time, it will generated a config.txt file in the user working directory. It will import the data config file into the use environment. This data will be used to change the column names of the imported dataset and change the name of the markers that is often incorrectly exported.

**Usage**

```
make_run_config(overwrite_config = FALSE, forcePath = NULL)
```

**Arguments**

overwrite_config	Boolean, if TRUE the config_drugsens.txt will be overwritten (default is FALSE)
forcePath	String, Define a custom path for the config file

**Value**

A dataframe/tibble.

**Examples**

```
# Generate config in temporary directory
make_run_config(forcePath = tempdir())
```

---

string_parsing	<i>Parse image filenames to extract metadata</i>
----------------	--

---

**Description**

This function will parse the data from the Image name and will return the metadata there contained  
The metadata will be then associated to the count file as well

**Usage**

```
string_parsing(.data)
```

**Arguments**

.data            dataframe with parsed metadata

**Value**

A dataframe/tibble.

**Examples**

```
# Basic example with sample data
input_data <- data.frame(
  Image = "B516_Ascites_2023-11-25_DOC2020-12-14_dms0_rep_Ecad_cCasp3_(series 01).tif"
)
test <- drugsens::string_parsing(input_data)
```

```
# Example with actual data processing
data_parsed <- string_parsing(input_data)
```

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